USE OF LICORICE EXTRACT IN COUNTERACTING AFLATOXICOSIS IN BROILERS

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ABSTRACT

This study was conducted at the Animal Production Farm/State Board for Agricultural Research over the period from 21/3/2002 to 27/4/2002. A total of 900 Fawbro broilers, three weeks - old were used to investigate the probable role of licorice extract in suppressing the deterimental effects of aflatoxicosis on productive performance of broilers, Chicks were randomly allocated to 6 treatments of 3 replicates. Birds in the first treatment (T1) were fed a basal diet and used as control group. Birds in T2 treatment fed a diet contaminated with aflatoxin, while birds in T3 treatment were fed a diet contaminated with aflatoxin and treated with mold killer. However, birds in T4, T5 and T6 treatments were fed a diet contaminated with aflatoxin and supplemented with licorice extract at the levels of 150, 300 and 450mg/kg of diet, respectively.

Inclusion of the aflatoxin in the diet resulted in a significant (p<0.05) decrease in body weight, weight gain, feed consumption, feed conversion efficiency, Productive Index, Economic Figure, dressing percentage with or without viscera; an enlarged liver, spleen, heart, and gizzard and increased in abdominal fat. When mold killer (T3) or licorice extract (T4, T5 and T6) were incorporated into the diet containing aflatoxin, they significantly (p<0.05) improved all these traits. However, licorice treatments surpasses T3 as regards live body weight, weight gain, Productive Index and Economic Figure. Furthermore, there was a trend that T6 recorded the best results in relation to productive characteristics included in this study in comparison with T3, T4 and T5 treatments.

It was concluded from this study that supplementation of licorice extract particularly at a level of 450 mg/kg to the diet contaminated with aflatoxin can depress the adverse effects of aflatoxicosis on productive performance of broiler chickens.

الدراجي وآخرون

مجلة العلوم الزراعية العراقية 36 (1) : 197 - 205 ، 2005

استخدام مستخلص عرق السوس لتقليل آثار التسمم يسموم الافلاتوكسين في فروج الحم

عماد الدين عباس العابي جاسم قاسم منابي حاتم عيسى الهيتي الهيأة العامة للبحوث الزراعية - وزارة الزراعة

حازم جبار الدراجبي قسم الشروة الحيوانية كلية الزراعة - جامعة بغداد

المستخلص

اجريت هذه الدراسة في حقل الانتاج الحيواني النابع لأهيأة العاسة البحوث الزراعية/وزارة الزراعة للمده مسن 2002/3/21 لغايسة 2002/4/27 وأستخدم فيها 900 فرخ فروج لحم فاوبرو بعمر ثلاثة اسابيغ لبحث الدور المحتمل لمستخلص عرق السوس فسبي تقليسل الآشار الضارة الناجمة عن تسمم العلف بسموم الافلاتوكسين. تو توزيع الافراخ عثروانياً على ثلاثة معاملات يتكون كل منها من ثلاثة مكررات وبواقسع 150 فرخا لكل معاملة . وتم تغذية الطيور في المعاملة الاولمي (٣١) على عليفة السيطرة وعنت كمجموعة قارنة . وغذيب الطيسور فسي المعاملة T2 على العليقة الملوثة بسموم الافلاتوكسين ، في حين ان طيور المعاملة T3 غنيت على عايقة ملوثة بسموم الافلاتوكسين ومعاملسسة بماده قائلة الفطريات Mold killer . من ناهية تانية ، فأن الطيور في المعاسلات 34 و 15 و 16 عذيت على عليقة ملوثة بسموم الافلائوكسيين ومضاف اليها مستخلص عرق السوس بمستويات 150 ر 300 و 450 مذهم/كفم علف على التوالي.

الشارت النقائج الى ان تلوث العلف بسموم الافلاتوكسين ادى الى تخفاض سعنوي (أ<0.05) في الوزن الحي للطير والزيادة الوزنيسة واستهلاك العلف وكفاءة التحويل الغذائي والدليل الانتاجي والمؤشر الاقتصادي ونسية التصافي سع او بدون الاحشاء الدالخلية وألمي زيادة معنويسة (أ<0.05) في الوزن النسبي للكيد والطحال والقانب والقانصة ودهن البطن. من ناحية ثانية ، عندما تمت اضافة المادة القاتلـــة العفــن (13) أو عرق السوس (T4 وT5 وT6) للعلف الملوث بسموم الافلاتوكسين قائها حسنت معنوياً (أ<0.05) جميع العنفسات العذكسورة بالمقارنسة مسع المعاملة T2. كما ان معاملات عرق السوس قد تفوقت على المعاملة 33 فيما يتعلق بوزن الجمع الحيي والزيادة الوزنيســة والدليــل الانقــاجي والمؤشر الاقتصادي . فضلاً على ذلك ، كان هذاك انجاء واضح بأن المعاملة T6 قد سجلت افضل النتائج نيما يتعلق بالصفات الانتاجيسة التسى شملتها الدراسة الحالية مقارنة بالمعاملات، T3 و T4 و T5.

يدتنتج من الدراسة الحالية بأن اضافة مستخلص عرق السوس خصوصاً بمستوى 450 ملغم/كغم للعلف الملونة بسموم الافلاتوكسين يمكن أن يحد من التأثيرات الضارة لهذه السموع في الأداء الإنتاجي لفروج اللحم.

^(*)Accepted on 1/12/2004 - Received on 12/9/2004

Introduction

The aflatoxin-producing fungi have been found in large variety of commodities. Condition favouring their growth and toxin production are: high moisture content, high temperature, insect damage, and the physical condition of the grain, weathering, mechanical handling and presence of cracked grains (13) .Reddy et al ., (15) reported that infestation of feeds with aflatoxins causes increased mortality, decreased growth rate and poor conversion in broiler chicken .However, aflatoxin was demonstrated to produce in broiler chicken a steatorrhea accompanied by a decrease in digestive enzymes elaborated by pancreas(14). Giambrone et al., (7) indicated that aflatoxin treatment resulted in increase in the susceptibility of broiler chickens to salmonellosis, aspergillosis, coccidiosis, Marek's disease. The immuno depressive action in aflatoxins was primarily on the cell-mediated immune system (7).

Aflatoxin is the common name for a group of structurally related compounds (aflatoxin B1, B2,G1, and G2) produced by fungi of the flavus - parasiticus group of the genus Aspergillus. This mycotoxin is potentially a threat to poultry health and production through contamination of poultry feeds (8). Al-Daraji (2) reported that experimentally induced aflatoxicosis resulted in significant deterioration in erythrocytes, leucocytes, thrombocytes, haemoglobin concentration, heterophil/ lymphocyte ratio, hematocrit and plasma uric acid, glucose, cholesterol, protein, calcium, phosphorus, GOT activity and alkaline phosphatase activity.

Licorice exerts numerous beneficial effects on the body, making licorice a valuable herb for treating a host of aliments. It can help reduce inflammation. It seems to prevent the breakdown of adrenal hormones such as cortisol (the body's primary stress - fighting adrenal hormone), making these hormones more available to the body and helps the body cope with stress (20). Licorice also appears to enhance immunity by boosting levels of interferon, a key immune system chemical that fights off attacking viruses (6). However, licorice is also known to exhibit many pharmacological actions, including estrogenic activity, anti - inflammation, allergic, antibacterial, antiviral, antihepatotoxic, fungicide, anticancer and anti - Trichomonas (12).

The present study was undertaken trial to suppress the effect of aflatoxicosis on productive performance of broiler chickens by using different levels of licorice extract. Licorice extract was

supplemented at levels of 150, 300, and 450 mg/kg to the diet of birds which was previously contaminated with aflatoxin.

Materials and Methods

This study was conducted at the Animal Production farm/State Board for Agricultural Research over the period from 21/3/2002 to 27/4/2002 A total of 900 Fawbro broiler, three weeks of age were used. Birds were fed starter diet during the third week of age (beginning date of experiment; 22.7% crude protein and 2867.4 kcai/kg of diet) and finisher diet (20.6% crude protein and 2922 kcal/kg of diet) until the marketing age (49 days of age). Chicks were randomly divided into 6 treated groups of 3 replicate per group, each replicate constitutes 50 chicks (150

chicks per treatment group).

Birds in the first treatment fed a commercial broiler ration and used as a control group (T1). The second treatment (T2) was fed a diet contaminated with aflatoxin, while birds in the third treatment (T3) were fed a diet contaminated with aflatoxin and treated with mold killer (Choong ang Biotech company, Korea). However, birds in fourth, fifth, and sixth treatments were fed a diet contaminated with aflatoxin and supplemented with licorice extract. Licorice extract was to the diet of birds supplemented throughout the total period of experiment at levels of 150 mg/kg (T4), 300 mg/kg (T5) and 450 mg/kg of diet (T6).

Aflatoxin used in the present study was aflatoxin B1 which obtained from the Department of Plant Protection, College of Agriculture, University of Baghdad. Aflatoxin was prepared and incorporated into basal diet by method previously reported (17). Aflatoxin was produced by growing Aspergillus flavus on rice. The moldy rice was dried and ground to a fine powder and analyzed spectrophotometrically for its total aflatoxin content by the method of Nabney and Nesbitt (10). The moldy rice then added to the yellow corn that involved in the basal diet. The final level of aflatoxin introduced to the birds was determined to be equal to 2 mg aflatoxin/kg of diet.

Productive characteristics measured in this study included: body weight, weight gain, feed consumption, feed conversion ratio, and mortality. However, Productive Index and Economic Figure determined according to Naji and Hana (11). At the end of experiment, 18 birds per each treatment (6 birds per each replicate) were sacrificed to determine dressing percentage with or without

viscera, and weights of visceral organs, viz. liver, heart, gizzard, spleen, in addition to

abdominal fat.

Significance of data was determined at the 5% level of probability by analysis of variance (ANOVA) using the Statistical Analysis of System (16). Significance of the differences between treatment means was determined by Duncan's multiple range test (16).

Results and Discussion Dietary aflatoxin (T2) significantly (P<0.05) depressed body weight, weight gain, feed consumption, feed conversion, Productive Index, Economic Figure, and liveability starting from the fourth week of age through the seventh week of age in comparison with control group (T1; Figures 1, 2, 3, 4 and Table 1). When mold killer (T3) or licorice extract (T4, T5 and T6) were added to the diet containing atlatoxin, they significantly (p<0.05) increased these traits. However, T4, T5, and T6 surpasses T3 treatment as regards live body weight and weight gain (Figures and 2). There were no significant differences between licorice treatments (T4, T5, and T6) and T3 throughout the experimental period with relation to feed efficiency and mortality conversion (Figure 4 and Table 1). Furthermore, there were no significant differences between licorice treatments and T3 treatment during 4th and 5th weeks of age, between T3, T4 and T5 treatments during the 6th week of age, and between T3 and T4 treatments during the 7th week of age in regard to

The addition of 450 mg/kg licorice extract to the diet containing aflatoxin restore the mean of live body weight, feed consumption, Productive Index Economic Figure to the control values (Figures 1, 2, 3 and Table 1).

feed consumption (Figure 3).

The results of this investigation clearly demonstrate that aflatoxicosis in broiler chickens can be influenced by supplementation the licorice extract to the contaminated diet .Increasing the licorice content of the diet to 450 mg/kg essentially negated the effects of aflatoxin. An obvious explanation to the protective effects of licorice is that licorice shows some anti-infective properties .In laboratory and animal studies, it has stopped or slowed down the growth of certain bacteria, fungi, and parasites. Several animal studies have also revealed a possibly strong antiviral and fungicide effects for true licorice (5) .In these studies, true licorice component that

belong to the isoflavonoid class of chemicals, appear to have several anti infective effects that include interference with oxygen utilization by infective organisms. Additionally, true licorice may have some ability to improve functioning of the immune system (1, 18). Newall et al. (12) reported that medicinal use of licorice in both Western and Eastern cultures dates back several thousands years. Licorice is know to exhibit many pharmacological actions. including anti-inflammatory (cortisol-like), antiviral, antibacterial, antifungal, anti-Trichomonas, antihepatotoxic and anti allergic activities. The plant reinforce the body's ability to withstand attack from virtually any kind of pathogen. However, if one is looking for a broadspectrum tonic to protect, maintain health, and heal injuries, there is no herb better than licorice root. Utsunomiya et al. (20) indicated that modern research on licorice reports many effects which are adrenal enhancing, analgesic, anti inflammatory, anti tumor, antiviral, immune protecting, liver antioxidant, fungicide, protecting and liver detoxifying. However, by functioning as anti-fungal agent, this herb destroys or prevent the growth of fungi.

Inclusion the aflatoxin in the diet (T2) resulted in a significant reduction in both dressing percentage with or without viscera compared with control group (T1; Table 1). However, the supplementation of mold killer (T3) and licorice extract (T4, T5, T6) to the aflatoxin – contaminated diet significantly improved these two traits in comparison with T2 treatment. T6 treatment surpasses other treatments in relation to dressing percentage with or without viscera and it restore the means of these two traits to the control values.

The effects of different treatments on the relative weight of certain organs are presented in Table 1. With incorporation of aflatoxin into the diet (T2), visceral organs such as liver, heart, gizzard and spleen significantly (p < 0.05) increased in comparison with control group (T1). However, administration of graded levels of licorice extract (T4, T5, T6) or mold killer (T3) resulted in significant reduction in the relative weight of these grants in the relative weight of these organs compared with T2 treatment. Furthermore, there was a trend that T6 recorded the lowest (p< 0.05) means regarding the relative weights of liver, heart, gizzard and spleen comparing with other treatments (T3, T4 and T5).

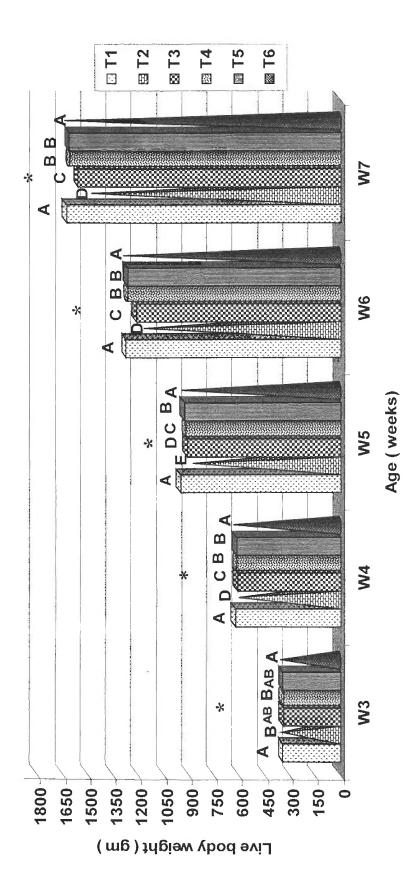


Figure 1. The effect of different levels of licorice extract on mean body weight of broiler fed a diet contaminated with aflatoxin.

T1= Birds fed a basal diet, T2= Birds fed diet contaminated with aflatoxin, T3=Birds fed diet contaminated with aflatoxin and treated with mold killer, T4= supplemented with licorice extract at level of 300 mg/kg, T6= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 450 Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 150 mg / kg, T5= Birds fed diet contaminated with aflatoxin and

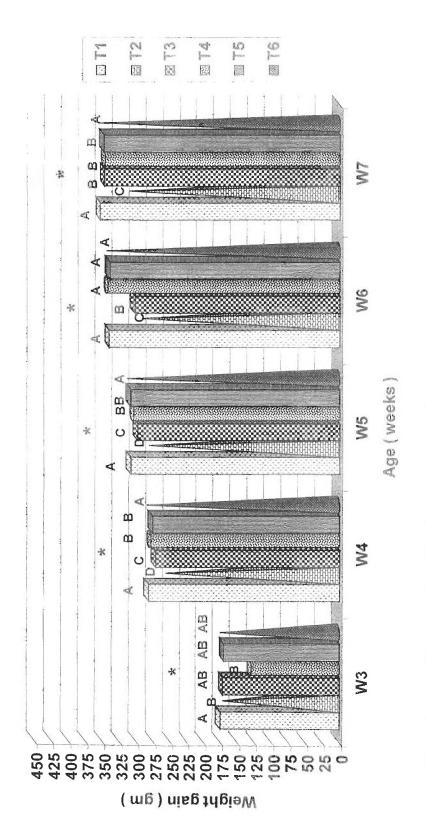


Figure 2. The effect of different levels of licorice extract on weight gain of broiler fed a diet contaminated with aflatoxin

supplemented with licorice extract at level of 300 mg / kg, T6= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 450 T1= Birds fed a basal diet, T2= Birds fed diet contaminated with aflatoxin, T3=Birds fed diet contaminated with aflatoxin and treated with mold killer, T4= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 150 mg/kg, T5= Birds fed diet contaminated with aflatoxin and

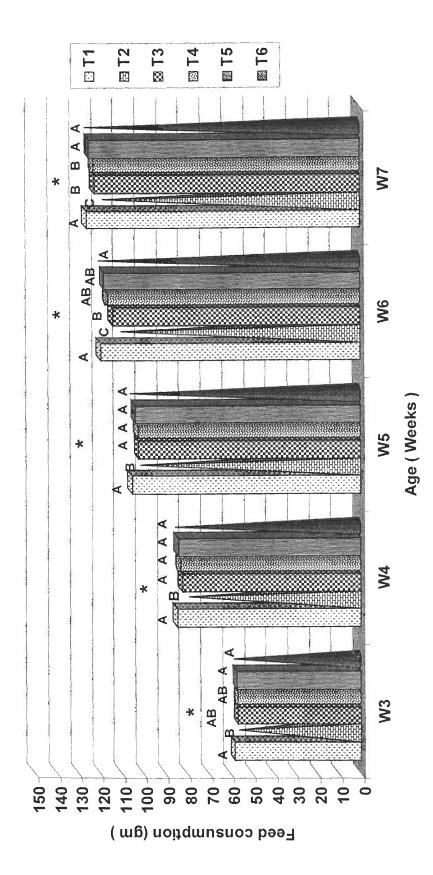
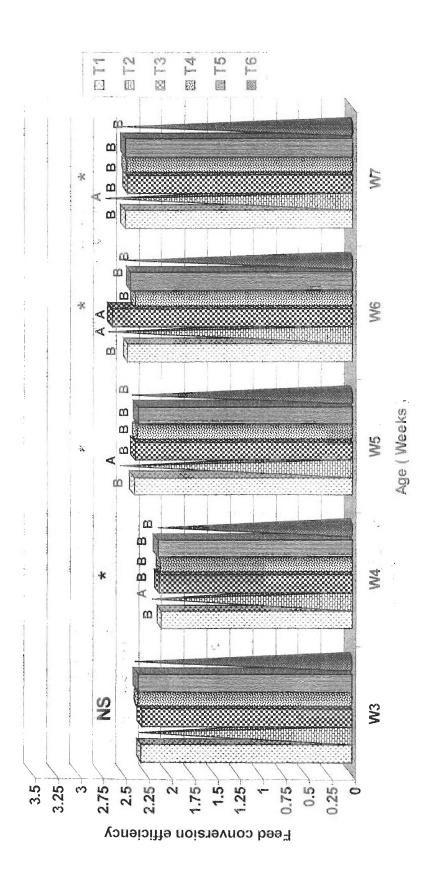


Figure 3. The effect of different levels of licorice extract on feed consumption of broiler fed a diet contaminated with aflatoxin.

supplemented with licorice extract at level of 300 mg / kg, T6= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 450 T1= Birds fed a basal diet, T2= Birds fed diet contaminated with aflatoxin, T3=Birds fed diet contaminated with aflatoxin and treated with mold killer, T4= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 150 mg/kg, T5= Birds fed diet contaminated with aflatoxin and



Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 150 mg / kg, T5= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 300 mg / kg, T6= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 450 mg T1= Birds fed a basal diet, T2= Birds fed diet contaminated with aflatoxin, T3=Birds fed diet contaminated with aflatoxin and treated with mold killer, T4=

Figure 4. The effect of different levels of licorice extract on feed conversion efficiency of broiler fed a diet contaminated with affatoxin

Table 1. The effect of different levels of licorice extract on certain productive performance and organ weights of broiler fed a diet contaminated with aflatoxin

Treatments Treatments					
T1	Т2	Т3	T4	Т5	Т6
A		_			AB
					$190.18 \pm$
-	20.00				12.71
		-			AB
					$191.01 \pm$
13.47	17.15				14.8
C	A				В
	$2.69 \pm$	$1.62 \pm$			$1.41 \pm$
1.39 ±0.00	0.11	0.08	0.08	0.06	0.05
A	C	В	В	В	A
$72.81 \pm$	$71.93 \pm$	$72.55 \pm$	$72.53 \pm$	$72.74 \pm$	$74.80 \pm$
4.69	7.33	6.28	6.19	5.63	4.82
A	C	В	В	В	A
65.14 ±	$63.92 \pm$	64.72 ±	64.77 ±	64.97 ±	65.1 ±
3.95	5.64	4.63	4.49	5.02	4.11
C	A	AB	C	В	С
1.04 ±	$1.11 \pm$	$1.09 \pm$	1.05 ±	$1.08 \pm$	$1.05 \pm$
0.09	0.01	0.01	0.01	0.08	0.01
D	A	В	В	В	C
$3.01 \pm$	$3.89 \pm$	$3.17 \pm$	$3.16 \pm$	$3.15 \pm$	$3.05 \pm$
0.08	0.16	0.07	0.09	0.09	0.07
D	A	В	В	В	C
$0.51 \pm$	$0.68 \pm$	$0.60 \pm$	$0.60 \pm$	$0.58 \pm$	$0.54 \pm$
0.05	0.01	0.08	0.09	0.08	0.06
D	A	В	В	В	C
$0.10 \pm$	$0.28 \pm$	$0.17 \pm$	$0.16 \pm$	$0.15 \pm$	$0.12 \pm$
0.003	0.009	0.005	0.005	0.004	0.03
D	A	В	В	В	C
$2.60 \pm$	$3.03 \pm$	$2.84 \pm$	$2.79 \pm$	$2.81 \pm$	2.66 ±
0.01	0.03	0.02	0.02	0.02	0.02
	$\begin{array}{c} T1 \\ A \\ 195.28 \pm \\ 12.21 \\ A \\ 196.04 \pm \\ 13.47 \\ C \\ 1.39 \pm 0.06 \\ \hline A \\ 72.81 \pm \\ 4.69 \\ \hline A \\ 65.14 \pm \\ 3.95 \\ \hline C \\ 1.04 \pm \\ 0.09 \\ \hline D \\ 3.01 \pm \\ 0.08 \\ \hline D \\ 0.51 \pm \\ 0.05 - \\ D \\ 0.10 \pm \\ 0.003 \\ \hline D \\ 2.60 \pm \\ \end{array}$	$\begin{array}{c cccc} T1 & T2 \\ \hline A & D \\ 195.28 \pm & 161.60 \pm \\ 12.21 & 15.69 \\ \hline A & D \\ 196.04 \pm & 162.14 \pm \\ 13.47 & 17.15 \\ \hline C & A \\ 1.39 \pm 0.06 & 2.69 \pm \\ \hline 0.11 & C \\ 72.81 \pm & 71.93 \pm \\ 4.69 & 7.33 \\ \hline A & C \\ 65.14 \pm & 63.92 \pm \\ 3.95 & 5.64 \\ \hline C & A \\ 1.04 \pm & 1.11 \pm \\ 0.09 & 0.01 \\ \hline D & A \\ 3.01 \pm & 3.89 \pm \\ 0.08 & 0.16 \\ \hline D & A \\ 0.51 \pm & 0.68 \pm \\ 0.05 & 0.01 \\ \hline D & A \\ 0.10 \pm & 0.28 \pm \\ 0.003 & 0.009 \\ \hline D & A \\ 3.03 \pm & 3.03 \pm \\ \hline \end{array}$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$

*T1= Birds fed a basal diet, T2= Birds fed diet contaminated with aflatoxin, T3=Birds fed diet contaminated with aflatoxin and treated with mold killer, T4=Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 150mg/kg, T5= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 300 mg/kg, T6= Birds fed diet contaminated with aflatoxin and supplemented with licorice extract at level of 450 mg/kg.

**Values in a row with different superscripts differ significantly (p < 0.05).

aflatoxicosis. Hepatic hypertrophy following toxin administration appeared to caused primarily by increased fat content in the organ (15). Mashaly et al.(9) reported that aflatoxin feeding at 50 µg/kg diet resulted in a significant decline in body and liver weights, rate of liver protein and RNA synthesis, and muscle RNA synthesis .Brown and Abrams (4) observed that a severe decline in plasma proteins on feeding aflatoxin B1 to chickens and ducklings was due to the suppression of liver protein synthesis as a consequence of mitochondrial injury and the lowered rate of ATP synthesis. Huff et al .(8) also noted that aflatoxin treatment significantly (p < 0.05) decreased body weight and

weight gain; increased the relative weight

Liver seemed to be the target organ

of the spleen, liver, proventriculus, gizzard, heart, and kidney; and induced hepatic hyperlipemia. Reddy et al. (15) found that with the increase of the level of aflatoxin, liver kidney, spleen, gizzard and pancreas showed an increase in weight with respective threshold doses of 0.50, 0.75, 1.50 and 4.0 ppm, while bursa of fabricius regressed at 1.25 ppm . Smith and Hamilton (19) demonstrated that graded doses of aflatoxin (1.25, 2.5, 5.0 and 10.0 ppm) incorporated into the feed of broiler chickens resulted in a decreased growth rate, an enlarged liver, spleen, and pancreas and a regressed bursa of fabricius. However, analysis of the liver showed that lipids accounted for 60 % of the dry weight increase (19).

The finding that licorice extract can the detrimental effects of aflatoxin on liver may be explained by that active component in licorice root are also used to help prevent and treat chronic hepatitis (liver inflammation). In one study of Japanese patients with hepatititis C those received intravenous treatment with glycyrrhizin for an average of 10 years were significantly less likely to develop liver cancer and cirrhosis (progressive liver failure; 21). In a second study of 57 patients with hepatitis C. glycyrrhizin (in dose ranging from 80 to 240 mg/day) significantly improved liver function after only one month. These effects diminished after glycyrrhizin treatment discontinued, however (3). Fujioka et al. (6) reported that licorice both protects the liver and promotes healing of this vital organ. The herb's anti - inflammatory properties help hepatitis-associated inflammation. Licorice also fights the virus and toxins commonly responsible for hepatitis, and supplies valuable antioxidant compounds that help maintain the overall health of liver and certain vital organs. However, glycyrrhizin may protect liver and other vital organs such as heart, spleen, and kidney from being damaged by oxidants. Too many oxidants can harm healthy cells and cause inflammation. Licorice root nutritionally supports the respiratory and gastrointestinal systems, liver, heart and spleen (22).

It was concluded from this study that dietary licorice extract especially at level of 450 mg/kg of diet can influence the severity of aflatoxicosis in broiler chickens and may be helpful in the control and prevention of this economically

important disease.

REFERENCES

1-Adam, L. 1997. *In vitro* antiviral activity of indigenous glycyrrhizin, licorice, glycyrrhizic acid (sigma) on Japanese encephalitis virus. J. Commun. Dis .29(2): 91–99.

2-Al-Daraji, H. J., I. A. Al-Ani, J. K. Minati and H. E. Al-Hiti. 2004. The use of different methods to suppress the effect of aflatoxicosis and it's influence on certain blood traits in broiler. Iraqi J. Agric. Sci. 35(2):103-112.

3-Arase, Y., K. Ikeda and N. Murashima. 1997. The long term efficacy of glycyrrhizin hepatitis C patients. Cancer

79 (8) :1494--1500.

4-Brown, J. M. M. and L. Abrams. 1965. Biochemical studies on aflatoxicosis. Onderstepoort J. Vet. Res. 32:119–146.

5-Duke, J. A. 1985. CRC Handbook of Medical Herbs. Boka Raton, Florida: CRC Press, USA.

6-Fujioka, T. Kondou and A. Fukuhara. 2003. Efficacy of glycyrrhizin suppository for treatment of chronic hepatitis C: a pilot study. Hepatol Res. 26 (1):103-117.

7-Giambrone, J. J., U. L. Diener, N. D. Davis, V. S. Panangala and F. J. Hoerr. 1985. Effect of aflatoxin on young turkeys and broiler chickens. Poultry Sci.

64: 1678-1684.

8-Huff, W. E., L. K. Kubena, R. B. Harvey, W. M. Hagler, Jr., S. P. Swanson, T. D. Phillips and C. R. Creger. 1986. Individual and combined effect of aflatoxin and deoxynivalenol (DON, Vomitoxin) in broiler chickens. Poultry Sci. 65:1291-1298.

9-Mashaly, R. I., M. H. Salem, Z. H. Mahmoud, S. A. El-Deeb, G. El-Sharawi and A. A. Ismail. 1986. Effect of aflatoxins on body-weight gains, and on protein and RNA synthesis in chickens .Indian J.Anim.Sci. 56(6):698-702.

10-Nabney, J. and B. F. Nesbitt. 1965. A spectrophotometric method of determining the aflatoxins. Analyst

90:155-160.

11-Naji, S. A. and A. G. Hana. 1999. Broiler Manual. Al-Hibba Bureau for Printing and Distribution. 1st ed. Baghdad, Iraq.

12-Newall, C. A., L. A. Anderson and J. D. Phillipson. 1996. Herbal Medicines: A Guide for Health-Care Professionals.

Pharmaceutical, London, U. K.

13-Obioha, W. I., H. M. Stahr and A. A. Kraft. 1986. Distribution and effects of aflatoxin in chicken tissues after feeding radiolabeled (¹⁴ C) aflatoxin B1. J. Food. Protec. 49(10):799–805.

14-Obsorna, D. J. and P. B. Hamilton. 1981. Decreased pancreatic digestive enzymes during aflatoxicosis. Poultry

Sci. 60:1818-1821.

15-Reddy, A. R., V. R. Reddy, P. V. Rao and B. Yadagiri. 1982. Effect of experimentally induced aflatoxicosis on the performance of commercial broiler chicks. Indian J. Anim. Sci. 52(6):405–410.

16-SAS. 1989. SAS User's Guide: Statistics (Version 5th ed). SAS. Inst. Inc.

Cary. N. C. USA.

17-Shotwell, O. L., C. W. Hesse and W. G. Sorenson. 1995. Production of aflatoxin on rice. Appl. Microbial. 14:425–428.

18-Shibata, S. A. 2000. Drug over the millennia: Pharmacognosy, chemistry,

and pharmacology of licorice. [Review]. Yakugaku Zasshi. 120(10):849-862.

19-Smith, J. W. and P. B. Hamilton. 1970. Aflatoxicosis in the broiler chicken.

Poultry Sci. 49:207-215.

20-Utsunomia, T., M. K. Kobayashi, D. N. Herndon, R. B. Pollard and F. Suzuki. 1999. Effects of glycyrrhizin, an active component of licorice root, on *Candida albicans* infection in thermally injured mice. Clin.Exp.Iramunol. 116: 291–298.

21-Van Rossum, T. G., A. G. Vulto, W. C. Hop, J. T. Brouwer, H. G. Niesters and S.

W. Schalm. 1999. Intravenous glycyrrhizin for the treatment of chronic hepatitis C: a double – blind, randomized, placebo–controlled phase 1/11. J. Gastroenterol Hepatol.14(11):1093-1099. 22-Vaya, J., P. A. Belinky, M. Aviram. 1997. Antioxidant constituents from licorice roots: isolation, structure elucidation and antioxidative capacity towards LDL oxidation. Free Radic. Biol. Med. 23(2):302–313.